Summary: Late infantile GM1 gangliosidosis is an extremely rare metabolic disorder with clinical features of seizure and progressive motor and mental retardation without facial dysmorphism or visceral organomegaly. We report the CT and MR imaging findings in one infant, which included abnormalities of the cerebral cortex, white matter, and deep nuclei.

GM1 gangliosidosis is a rare lysosomal storage disease characterized by a deficiency in the activity of lysosomal β-galactosidase, which results in increased accumulation of GM1 ganglioside and asialo-GA1 in the brain and of oligosaccharide in the visceral organs (1, 2). Patients with infantile GM1 gangliosidosis usually have dysmorphic facial features, motor and mental retardation, seizures, and hepatosplenomegaly (2). The late infantile form of this disorder is extremely rare and has no clinical features of facial dysmorphism or visceral organomegaly (1).

Case Report

An 11-month-old girl, the product of an uncomplicated full-term pregnancy, presented with complex febrile convulsion during an episode of upper respiratory infection. Physical examination at first admission showed a hypotonic infant with macrocephaly (>97 percentile), normal facial features, and psychomotor retardation. Hyperactive reflexes and augmented startle response to noise were present. The abdomen was soft without hepatosplenomegaly. Examination of the eyes revealed a normal clear lens, dilatation of the pupil without light reflex in the right eye, and mild dilatation of the pupil with sluggish light reflex in the left eye. Fundoscopy showed no cherry-red spots at the maculae. Deficient β-galactosidase activity and normal hexosaminidase, and glucosidase activity on lysosomal enzyme assay in cultured skin fibroblasts were documented at the patient's second admission 3 months later for clonic-tonic seizure and progressive motor and mental retardation between 1 and 5 years of age.

Seizures are common, and spastic tetraplegia develops, with cerebellar and extrapyramidal signs. Decerebrate rigidity follows, and death occurs between 3 and 10 years of age, usually precipitated by recurrent bronchopneumonia. Dysmorphic facial features, hepatosplenomegaly, corneal changes, and bony abnormalities are usually lacking, and, when present, are due to a mild degree or absence of oligosaccharide accumulation. Cerebral GM1 ganglioside storage is also less severe in type 2 than in type 1 disease (1, 2).

Discussion

GM1 gangliosidosis is a rare inborn error of metabolism caused by a deficiency of β-galactosidase activity resulting in failure of catalyzing cleavage of terminal β-link galactose from substrates, such as GM1 ganglioside, asialo-GM1, lactosylceramide, galactose-containing oligosaccharides, and mucopolysaccharides. The abnormally accumulated substances in the lysosomes of the affected cells in the CNS are mainly GM1 ganglioside and its asialo derivative GA1 as well as other minor glycolipids and glycopeptides. Visceral storage with oligosaccharides is variable and may cause organomegaly.

Three types of GM1 gangliosidosis, classified by age of onset, have been reported (1, 3, 4). Infantile GM1 gangliosidosis (type 1) is the most common and severe form, with clinical features of hypotonia, failure to thrive in the neonatal period, and clonic-tonic seizure activity. Coarse facial features, frontal bossing, dysostosis multiplex, hepatosplenomegaly, and hazy cornea may be present or become apparent in the first year of life (2). The late infantile or juvenile form (type 2) begins with progressive mental and motor retardation between 1 and 5 years of age. Seizures are common, and spastic tetraplegia develops, with cerebellar and extrapyramidal signs. Decerebrate rigidity follows, and death occurs between 3 and 10 years of age, usually precipitated by recurrent bronchopneumonia. Dysmorphic facial features, hepatosplenomegaly, corneal changes, and bony abnormalities are usually lacking, and, when present, are due to a mild degree or absence of oligosaccharide accumulation. Cerebral GM1 ganglioside storage is also less severe in type 2 than in type 1 disease (1, 2).
Children or adults with chronic, or type 3, GM1 gangliosidoses may have a slowly progressive disorder in which dystonia, dysarthria, ataxia, myoclonus gait disorders, and extrapyramidal signs occur (4). Bony changes are minimal. Cherry-red spots at the maculae, which are found in about half the patients with type 1 GM1 gangliosidoses, are not seen in patients with type 2 and type 3 disease.

The disorder can be diagnosed in several ways, including lysosomal enzyme assay of low β-galactosidase activity in peripheral leukocytes or cultured skin fibroblasts, detection of abnormal urinary oligosaccharide excretion, and rectal biopsy (2). Prenatal diagnosis by measurement of enzyme activity in amniotic fluid and cultivated amniotic fluid cells has also been established (5).

Neuropathologic reports on GM1 gangliosidoses have shown diffuse neuronal storage with ballooning of neuronal cytoplasm in the cerebral and cerebellar cortex, basal ganglia, brain stem, spinal cord, and dorsal root ganglion in type 1 and type 2 forms, and a tendency for selective storage in the basal ganglia with neuronal loss and gliosis in type 3 disease (6, 7). The cerebral white matter is gliotic and there is loss of myelin in type 1 but not in types 2 and 3 disease.

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Neuromaging findings in patients with type 1 GM1 gangliosidoses have been reported only in a few cases. In one case, initial thalamic hyperdensity was found on CT scans and hypointense signal of the thalamus was seen on T2-weighted MR images at a later stage (3). Persistent delay in white matter myelination on serial MR studies has been described in another case (8). In type 3 GM1 gangliosidoses, the symmetrical abnormal signal intensities were more selectively seen in the caudate nucleus and putamen on T2-weighted MR images (9). To our knowledge, neuromaging findings in type 2 GM1 gangliosidoses have not been reported previously.

The CT and MR findings in our case are identical to those reported in patients with Tay-Sachs disease (GM2 gangliosidoses). This may be understood because the structure of gangliosides GM1 and GM2 differs only in the terminal N-acetylgalactosamine, and they are stored together with cholesterol and phospholipid; the neuronal staining reactions and ultrastructure are identical (7). The thalamic hyperdensity seen on CT scans in Tay-Sachs and Krabbe lysosomal storage disorders has been presumed to be due to calcification. Calcium deposition in the thalamus can also explain the hyperintense T1 signal and hypointense T2 signal on MR images, although frank calcium deposition has not been reported pathologically (10). Other investigators have suggested that secondary lipofuscinosis, resulting in deposits of pigmented degradation products, occurs in the thalamus and cause this appearance in GM2 gangliosidoses (11, 12). In our case, the CT study showed not only thalamic hyperdensity but also hypointensity of the basal ganglia, which were almost isointense with adjacent white matter (Fig 1A). The MR study verified the CT findings of abnormal thalamic attenuation but not the hypointensity of the basal ganglia. MR is the only imaging technique that is capable of demonstrating...
the arrested myelination of the cerebral white matter in serial examinations (Fig 1C and D).

Conclusion

Although the neuroimaging findings of Gm1 gangliosidoses are unique in our case, these findings could also be identified in Gm2 gangliosidoses and, to some extent, in late-stage Canavan disease. Patients with late-stage Canavan disease may have dense thalami on T1-weighted images, as well as white matter necrosis, resulting in cavitation, and brain stem and cerebellar atrophy, which are not observed in Gm1. Other neurometabolic diseases that often manifest with macrocephaly include mucopolysaccharidosis and Alexander disease. These disorders have distinct neuroimaging features and they should be distinguished from Gm1. A definite diagnosis of this rare disorder can only be made by obtaining lysosomal enzyme assay results of deficient \( \beta \)-galactosidase and normal hexosaminidase.

References


